# Investigation on Tularemia in potential reservoirs' of Anatolia\*

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**Summary:** Tularemia is a zoonotic infection caused by *Francisella tularensis* and has gained renewed importance since there has been a recent increase in the number of human cases in several countries across the world. In this study the existence of *F. tularensis* in rodents and in water was investigated by culture and PCR techniques. Also, *F. tularensis* specific antibodies were investigated by serological methods in sheep. At the end of the study, *F. tularensis* was isolated from one water sample by culture and PCR techniques, on the other hand, 27.6% seropositivity was detected in the blood samples of sheep.

Key words: Francisella tularensis, rodent, sheep, water, zoonoses.

## Anadolu'nun potansiyel rezervuarlarında Tulareminin araştırılması

Özet: Tularemi, *Francisella tularensis* tarafından oluşturulan zoonotik bir enfeksiyondur ve dünya çapında bazı ülkelerde insan vakalarının artmasından dolayı yeniden önem kazanmıştır. Bu çalışmada rodentlerde ve suda *F. tularensis*'in varlığı kültür ve PCR teknikleriyle araştırılmıştır. Bunun yanı sıra, serolojik metodlarla koyunlarda *F. tularensis* spesifik antikorlarının varlığı araştırılmıştır. Çalışmanın sonunda, bir adet su numunesinden *F. tularensis* izole edilmiştir ve diğer taraftan koyun kan numunelerinde %27.6 seropozitiflik tespit edilmiştir.

Anahtar sözcükler: Francisella tularensis, koyun, rodent, su, zoonoz.

## Introduction

*Francisella tularensis* is classified as a small pleomorphic Gram-negative coccobacillus. Its isolation dates back to 1911 from ground squirrels found dying of a plague-like illness in Tulare County, CA, USA. Initially, this agent called *Bacterium tularense*, then eventually classified to a new genus named *Francisella* inspired from the name of the man who pioneered research on the organism (11).

The risk of laboratory-acquired infection with of *F*. *tularensis* is an important issue that has to be dealt with extreme care. While working with live bacteria in the laboratory, biosecurity level III precautions should be met. In addition, while working with suspected samples also, biosecurity level II precautions should be provided.

The number of residential areas endemic for tularemia has been increasing rapidly in Turkey and there are many such today. Besides, the reservoirs of the disease are still unknown. Consistent with the changes in ecologic balances, the number of tularemia cases in Turkey has been increasing. It is thought that, the increase in the rodent population after rainy seasons could be responsible for the increase in tularemia cases. Transmission occurs usually via contact with infected animals, contaminated water and food, reservoir animals, wild animals and pets and by tick biting. Sheep are more sensitive to tularemia than other animals, but the disease can also be seen in other animals such as cats, dogs, pigs and horses. Rodents are generally believed to be the natural reservoir of the bacterium.

Postmortem examination is important when there is no clear information about the clinical symptoms of the tularemia in animals. During necropsy, enlarged spleen and white necrotic foci in the liver are noticed. When these findings are available, then it can be concluded that an animal sensitive to tularemia is in question (8). The aim of this study was to investigate the existence of F. *tularensis* in rodents and in water by culture and PCR techniques. In addition, spesific antibodies for F. *tularensis* were investigated by serological methods in sheep.

### Materials and Methods

*Sampling:* A total of 445 samples, including rodents, sheep blood sera were collected from 8 different residental areas in Turkey and 4 water samples from

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Ankara/Beypazarı. Details of the sampling period and the distribution of serum samples collected from these regions of Turkey is represented in Table 1 and Figure 1.

Table 1. Distribution of examined rodent, sheep blood serum and water samples according to the residental area. Tablo 1. Incelenen rodent, koyun kanı ve su materyallerinin

verlesim birimlerine göre dağılımı.

Region	Rodent	Sheep Blood Serum	Water	
Van	3	-	-	
Ankara-Beypazarı	3	101	4	
Ankara-Altındağ	-	46	-	
Bursa M. Kemalpaşa	10	159	-	
Zonguldak Kurtköy	3	20	-	
Ankara- Bala	7	75	-	
Sivas Gürün	9	-	-	
Yozgat	5	-	-	
Total	40	401	4	

*Strains and Reagents:* For isolation of *Francisella tularensis* from rodent organs and from water, antibiotic Francis medium was used. The medium was composed of brain heart infusion agar, dextrose (Difco) 1%, L-Cystein (Aldrich) 0.1%, sheep blood 8-9%, antibiotics (penicillin G 1ml/100 IU, Cyclohexamide L/100mg, Polymyxin B 1/8x10<sup>4</sup>) and Helicobacter supplement (8).

*Francisella tularensis* subsp. *holarctica* (NCTC 10857) strain was used as the positive control in the identification of the strains isolated from cultures, for testing the media and for molecular examinations of rodent and water samples. *Francisella tularensis* subsp. *tularensis* (SCHU S4), another positive control strain, was used for the subspecies determination in molecular examination. A tularenia antigen stained with safranine O which was prepared from *Francisella tularensis* subsp. *holarctica* (NCTC 10857) vaccine strain was used in the microagglutination test. *Francisella tularensis* antiserum was used as positive control (at 1/160 titer) in lam agglutination test, microagglutination test and in verifying the replicating colonies.

Primers used for determining the species and subspecies of the agents isolated are listed in Table 2 and 3.

Isolation: Isolation of the agents from the rodent and water samples: In order to detect the rodents that were bacteremic with *F. tularensis*, liver and spleen samples from 40 rodents were inoculated into the Francis medium supplemented with antibiotics and defibrinated sheep blood. After inoculation processes, dishes were incubated at  $37^{\circ}$ C for 10 days under 10% CO<sub>2</sub> environment. Colony morphology, physical features and development times of forming colonies were evaluated (5,8). In order to isolate the agent from water, samples



Figure. 1. Distribution of serum samples collected from various regions of Turkey. Şekil 1. Türkiye'nin farklı bölgelerinden toplanan serum örneklerinin dağılımı.

Table 2. The nucleotide composition of primers, the specific gene areas they amplify, and the length of the PCR products.
Tablo 2. Primerlerin baz dizileri bağlandıkları spesifik gen bölgeleri, PCR ürünlerinin uzunlukları.

Primer Name	Primer Sequence	Target gene	Length of product	References	
Tul 4-435 (F)	5`-GCT GTA TCA TCA TTT AAT AAA CTG CTG-3'	tul 4	420 hr	[16]	
Tul 4-863 (R)	5`-TTG GGA AGC TTG TAT CAT GGC ACT-3'	<i>IUI</i> 4	420 bp	[16]	

Table 3. Primer base groups and specific genes they amplify, PCR product lengths for the sub-species examinations used. Tablo 3. Alt tür tayininde kullanılan primerlerin baz dizileri bağlandıkları spesifik gen bölgeleri, PCR ürünlerinin uzunlukları.

Primer Name	Primer Sequence	Target gene	Length of product	References	
RD1 (F)	5'-TTT ATA TAG GTA AAT GTT TTA CCT GTA CCA-3'		024 hr	[2]	
RD1 (R)	5'-GCC GAG TTT GAT GCT GAA AA-3'	rd1	924 bp	[3]	

taken from 4 different sources at a volume of 500 ml were used. The water samples were filtered through a sterile cellulose acetate membrane filter (0.20  $\mu$ m pore diameter). After filtration process, the filter was placed on the antibiotic and defibrinated sheep-blood-supplemented Francis medium with the inoculated side upwards (16). Colonies growing at the end of the incubation period were subjected to lam agglutination test with *F. tularensis* positive immune serum and confirmation of the isolates were performed with molecular techniques.

Microagglutination test: Francisella tularensis specific agglutinins were examined by using microagglutination test (MAT) with antigen (0.004% including spleen-O) prepared with F. tularensis strains (NCTC 10857) (5). First, 40 µl saline buffer was put into the first well of the U based plate and 25 µl of saline put into the other 6 wells of the plate. 25  $\mu$ l of positive serum against tularemia was put into the 8th well (positive control 1/160 titer) and 10 µl of test serum (TS) into the first well. Next, 25 µl was transferred from the first well to the other. This dilution process performed till the 6th dilution. Last, 25 µl of dilute was taken out from the 6th well and dilution was completed. The 7th well did not receive any serum (negative control). Then all of the wells received 25 µl of stained antigen, thus 1/10-1/640 dilutions were prepared. Test plate was put into a humidified box and placed into the incubator. After incubation overnight at 37°C, evaluation of the test was performed according to the positive, negative and antigen controls (4,5).

*PCR analysis*: For the purpose of molecular diagnosis, conventional PCR (Polymerase Chain Reaction) was used in water samples, rodent liver and spleen samples and to confirm isolates. Extraction, amplification, electrophoresis and visualization operations were performed respectively (8).

In order to confirm *F. tularensis* with molecular techniques; a two stage PCR method was used to define

the species and subspecies of the samples. First, the *tul4* gene encoding a 17 kDa membrane protein was studied in order to direct the detection of genetic material from rodent samples and verify the culture-positive water samples. The band seen at 420 bp indicated *Francisella* spp. PCR (+) (13). Next, for differentiation of subspecies of *F. tularensis*, The Region of Difference 1 (RD1) target gene sequence was amplified from the culture positive water samples (3,8,13).

The serological and molecular studies of this research has been performed at the BSL-2 laboratory of Turkish Public Health Institution Administrative whereas isolation studies accomplished at the BSL-3 laboratory of the same Administrative.

## Results

Bacterial isolation from rodent and water samples: In total, 40 rodent samples were collected from 6 residential areas including Ankara Beypazarı, Sivas Gürün, Yozgat, Van and Bursa. In addition, 4 water samples were collected from Beypazarı (Ankara) to perform culture analysis. While no positive samples were detected by culture or PCR tests from rodents, one positive sample was detected from the 4 water samples and this was confirmed by PCR. Suspected colonies growing after incubation in culture media from water sample and isolation in Francis media were subjected to lam agglutination test with tularemia immune serum and this verified the presence of *F. tularensis* spp.

Serological Results: Serologic microagglutination tests of 401 blood sera collected from 5 different regions (Ankara Bala, Beypazarı, Altındağ, Zonguldak Kurtköy, Bursa Mustafa Kemal Paşa) revealed the following results: Of the 401 sera, in 156 (38.9%) the antibody titer was between 1/10 and 1/640 while 111 (27.6%) exhibited 1/20 antibody titer which was also interpreted as a positive tularemia titer (Table 4).

PCR Results: Francisella spp. specific DNA could not be detected in rodent samples. The results were in

Table 4. Results of microagglutination tests in sheep blood sera. Tablo 4. Koyun kan serumlarının mikroaglutinasyon test sonuçları.

	Number of materialsPositive serum samples		Titers					_		
Region			1/10	1/20	1/40	1/80	1/160	1/320	1/640	Percentage
Ankara/ Altındağ	46	17		13	4					36.95
Ankara/ Bala	75	63		19	18	18	6	1	1	84
Ankara/ Beypazarı	101	17	12	5						16.83
Zonguldak/ Kurtköy	20	1	1							5
Bursa/ M.Kemal Paşa	159	58	32	21	5					36.47
Total	401	156	45	58	27	18	6	1	1	38.9
Tularemia titer Percentage of sheep blood sera		38.9	28.85	37.18	17.30	11.54	3.85	0.64	0.64	

agreement with the culture findings. From the 4 water samples, 1 sample was positive in culture, gender-specific (tul4), after the primers went through PCR 420 bp of the band was seen so it indicated the presence of a *F. tularensis* spp. (Fig 2). a band at the 924bp area verifying that agent was *F. tularensis subspecies holarctica*. Thus, culture isolation of water sample was confirmed with PCR (Fig 3).

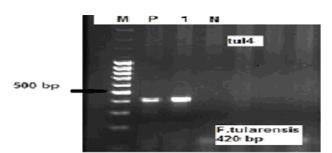


Figure 2. The image after PCR on the culture positive water samples of the gained agarose gel.

Resim 2. Kültür pozitif su örneğinin PCR sonrası elde edilen agaroz jel görüntüsü.

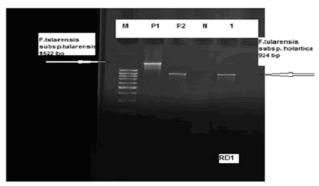


Fig 3. The image showing agarose gel after PCR for F.tularensis sub-species identification.

Resim 3. PCR sonrası F.tularensis alttür ayrımını gösteren agaroz jel görüntüsü.

# **Discussion and Conclusion**

The natural reservoirs for tularemia are mostly rodents. The disease is fatal for these animals when they are infected. Sheep, goats, cattle and other farm animals in addition to cats, dogs, horses and other pets are random hosts for the bacterium. In this study, existence of *F. tularensis* in sheep and other potential reservoirs was investigated. Furthermore, water samples were examined for isolation purposes.

A study by Arata et al. (1) on a variety of animals in Iran, analyzed 3548 spleen samples collected from 4600 wild animals potentially associated with tularemia. No *F*. *tularensis* isolates were obtained from any sample, but the agglutination tests of sera from 100 sheep, 100 cattle and 39 wild mammals, revealed a positive reaction in 3 cattle, 8 sheep and a hedgehog. Özsan et al. (12), reported no isolation of *F*. *tularensis* from 1379 wild

animal samples. However, when they injected liver and spleen suspensions prepared from wild animals into guinea pigs, they observed seropositivity between 1/40-1/80 by agglutination tests. In the light of these studies, they concluded that these animals might be the potential hosts for *F. tularensis*.

In our study, all isolation and PCR analyses were negative. In total, 40 spleen and liver samples collected from rodents in various locations were analyzed. While collecting rodent samples, it was attempted to sample from locations where tularemia in humans has been reported. During the study, the information that the number of rodents had increased was received from residents. In agreement with previous reports, we also had difficulties to directly isolate *F. tularensis* from the animal samples (15).

The first water-based tularemia isolate was made by Karpoff Antoroff (9) in Russia. Somewhat later, Hussein Oz (6) performed the first isolation from Turkey. Even though it is unclear how the bacterium contaminates water, it is speculated that the water is contaminated with the agent from rodent carcasses or other contaminated materials (2). Since the pollution in water is sudden and highly diluted and due to late sampling time, the chance of agent isolation is low from water. During our study, information of an epidemic from Ankara Beypazarı location was obtained. According to this information, 4 water samples were taken from this location and isolation and conventional PCR studies were conducted. At the end of the tests, one water sample of the 4 was found to be positive by culture isolation. This isolate was confirmed by conventional PCR technique revealing that the agent was F. tularensis subsp. holarctica. This result is important and demonstrates that the agent can be spread by water.

In a study conducted in United States, Jellison and Kohls (7) detected an antibody titer above 1/20 in 25% of sera from 148 lambs and accepted this titer as positive for *F. tularensis*. The researchers conducted another study involving 283 ewes returning from grassland and found 28.3% seropositivity.

In the present study, in order to investigate tularemia in sheep serologically, blood samples were collected from sheep in the locations where disease had been observed in humans. Samples were collected from 401 sheep from Ankara-Altındağ, Ankara-Beypazarı, Ankara-Bala, Bursa-Mustafakemalpaşa, Zonguldak-Kurtköy districts. Of the 401 sera, 156 (38.9%) had antibody titers between 1/10 and 1/640 while 111 (27.6%) exhibited 1/20 antibody titer which were also accepted as indicative of tularemia. O'Toole et al. (10) studied the existence of F. tularensis in 3 different local herds of sheep between 1997 and 2007. The report describes 4 periods of tularemia in these herds. Herd owners reported that ticks were unusually numerous and

often present on the sheep during outbreaks. Tularemia presented as late-term abortions and death in lambs and, to a lesser extent, in ewes. The diagnosis was corroborated by bacterial isolation and, in individual cases, by serology, and PCR detection of F. tularensis. These studies represents an example that indicate the association of tularemia between humans and sheep. Also, it indicates that the bacterium can be transmitted to sheep from ticks. Parallel to these findings, during our study in Ankara Beypazarı region Macunköy countryside, an epidemic of tularemia broke out in humans. From the area 2 rodents, 4 water samples from a public use, and 90 blood samples from sheep were taken. The agent was not isolated from the rodents, but on the other hand, while there was a positivity in water at this location, low antibody titers were detected in sheep that were drinking the water of the location. These results suggested that the possible transmission routes of the agent might be ticks rather than water.

Among the selected areas, a positive tularemia titer was observed in of 84% of the sera was found in Bala region as highest. This percentage is the highest seropositivity ratio in lambs ever seen in Turkey. From this perspective, our study is thought to be important and therefore represents an indication of the widespread distribution of sheep to the disease, detection of disease level and identifies the association of tularemia between human and sheep. Şeyda (14), analyzed 1600 sheep sera from the Kars region by microagglutination tests and detected 1/20 or higher antibody titers in 50 sheep.

In this study, the existence of *F. tularensis* was investigated by culture and PCR in rodents and water and by serological methods in sheep in locations selected in parallel with human tularemia cases.

Since we have studied several aspects of the tularemia epidemiology including several residential areas, this study is the first example in veterinary medicine in Turkey. In addition, serological results obtained are among the highest agglutination titers observed in sheep anywhere in the world. However, it is not possible to explain how the disease is transmitted in the areas with high titers, without identifying the reservoirs and potential vectors of the bacterium. The isolation of the agent has not been successful in studies of rodents. In rodents, after the entry of the organism into the body, mortality is likely to occur in a short time. In wildlife, it is unlikely to find a dead rodent, and in materials from live and healthy rodents caught, the chance of isolation is very low as reported from previous studies. It is not possible to suggest that rodents are not potential reservoirs because there are no isolates. Waterassociated positivity was found in areas where tularemia outbreaks took place. In this extensive study, the importance of these topics were emphasized by isolation trials from rodent and water samples, PCR tests and in

sheep by serological findings. A parallelism was observed in findings in isolation and PCR carried out. Therefore, the determination of the existence of the agent during a short time in the samples to be studied with PCR is very important.

In the studies to be carried out in future, periodical checks of samples from selected pilot locations, routine rodent control activities, and examination of humans seems to be important. Furthermore, considering waterassociated transmission, concurrent studies in humans, animals and water will be very important in the epidemiological evaluation of tularemia. At the end of the study, since paralel findings was observed by both isolation and PCR methods, it can be concluded that it would be important for rapid diagnosis of the existence of agent in samples to be examined by PCR. In addition, serological findings exhibit the highest agglutination titers in sheep ever known worldwide researches. With these findings, it is aimed to emphasize the importance of tularemia which is emerging as a popular zoonotic disease.

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